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The genetics of feto-placental development: A study of acid phosphatase locus I and adenosine deaminase polymorphisms in a consecutive series of newborn infants

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Published: 3 September 2008

Received: 14 January 2008

Reproductive Biology and Endocrinology 2008, **6**:38 doi:10.1186/1477-7827-6-38

Accepted: 3 September 2008

This article is available from: <http://www.rbej.com/content/6/1/38>

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Abstract

Background: Acid phosphatase locus I and adenosine deaminase locus I polymorphisms show cooperative effects on glucose metabolism and immunological functions. The recent observation of cooperation between the two systems on susceptibility to repeated spontaneous miscarriage prompted us to search for possible interactional effects between these genes and the correlation between birth weight and placental weight. Deviation from a balanced development of the feto-placental unit has been found to be associated with perinatal morbidity and mortality and with cardiovascular diseases in adulthood.

Methods: We examined 400 consecutive newborns from the Caucasian population of Rome. Birth weight, placental weight, and gestational length were registered. Acid phosphatase locus I and adenosine deaminase locus I phenotypes were determined by starch gel electrophoresis and correlation analysis was performed by SPSS programs. Informed verbal consent to participate in the study was obtained from the mothers.

Results: Highly significant differences in birth weight-placental weight correlations were observed among acid phosphatase locus I phenotypes ($p = 0.005$). The correlation between birth weight and placental weight was markedly elevated in subjects carrying acid phosphatase locus I phenotypes with medium-low F isoform concentration (A, CA and CB phenotypes) compared to those carrying acid phosphatase locus I phenotypes with medium-high F isoform concentration (BA and B phenotypes) ($p = 0.002$). Environmental and developmental variables were found to exert a significant effect on birth weight-placental weight correlation in subjects with medium-high F isoform concentrations, but only a marginal effect was observed in those with medium-low F isoform concentrations. The correlation between birth weight and placental weight is higher among carriers of the adenosine deaminase locus I allele*2, which is associated with low activity, than in homozygous adenosine deaminase locus I phenotype I carriers ($p = 0.04$). The two systems show a cooperative effect on the correlation between birth weight and placental weight: the highest value is observed in newborns carrying adenosine deaminase locus I allele*2 and acid phosphatase locus I phenotypes with medium-low F isoform concentration ($p = 0.005$).

Conclusion: These data suggest that zygotes with low adenosine deaminase locus I activity and low F activity may experience the most favourable intrauterine conditions for a balanced development of the feto-placental unit.

Background

We have recently described a cooperative interaction between ACP₁ (acid phosphatase locus 1) and ADA₁ (adenosine deaminase locus 1) genetic polymorphisms concerning their effects on the susceptibility to spontaneous primary repeated miscarriages: women carrying the ADA₁*2 and ACP₁*C alleles show the lowest susceptibility to repeated miscarriages [1]. Both systems share important effects on glucose metabolism and immunological function. These observations prompted us to search for a possible cooperative interaction between the two systems regarding their effects on developmental parameters during intrauterine life.

It is likely that a balanced growth of the two portions of the fetoplacental unit (i.e. without the prevalence of placental on the fetal part or vice versa) represents an advantage for fetal development. The birth weight/placental weight ratio (BW/PW) has been found to be correlated with perinatal morbidity and mortality and with cardiovascular disease in adulthood [2,3]. There is evidence that in addition to maternal factors and socioeconomic status, genetic factors also influence the ratio BW/PW [3-5]. In a recent note we have proposed the correlation between BW and PW as an index of balanced development of the fetoplacental unit and have shown that this correlation is influenced by ACP₁ phenotype [6].

The ACP₁ genetic polymorphism

ACP₁ also called low molecular weight phospho-protein tyrosine phosphatase (LMPTP) is an enzyme controlled by a locus on chromosome 2 showing three common alleles: ACP₁*A, ACP₁*B, ACP₁*C. These three alleles are associated with different enzymatic activities [7]. Activity of ACP₁ phenotypes are in the following order: A < BA < B = CA < CB < C [8].

Each allele at the ACP₁ locus encodes two isoforms, called F (fast) and S (slow) [9,10]. ACP₁ B and BA show a medium-high F isoform activity while CB, A, CA and C phenotypes show a medium-low F isoform activity. ACP₁ C, CA and CB show a much higher activity of S isoform as compared to other ACP₁ phenotypes [7,10].

Two important functions have been suggested for ACP₁: flavin-mono-nucleotide phosphatase activity and tyrosine phosphatase activity [11-13]. Catalysing the conversion of flavin-mononucleotide (FMN) to riboflavin, ACP₁ may have a role in regulating the cellular concentration of flavin-adenine-dinucleotide (FAD), flavo-enzyme activity and energy metabolism. As a phosphotyrosine phosphatase, the enzyme may have an important role in cellular growth regulation and in modulation of glycolytic rate through the control of receptor activities and of band 3 protein phosphorylation status [[12,14] and [15]].

Recently it has been shown that ACP₁ specifically dephosphorylates the negative regulatory Tyr-292 of ZAP-70, thereby counteracting inactivation of ZAP-70. The ZAP-70 protein-tyrosine kinase plays a central role in signalling from the T cell receptor. Thus, these results indicate that ACP₁ strengthens T cell receptor signalling [16].

The ADA₁ genetic polymorphism

ADA₁ is a polymorphic enzyme present in all mammalian tissues [17]. It is controlled by a locus with two codominant alleles ADA*1 and ADA*2 located on the long arm of chromosome 20. The corresponding three common ADA₁ phenotypes have different enzymatic activities: the ADA₁1 phenotype is 15% more active than the ADA₁2/1 phenotype and 30% more active than the ADA₁2 phenotype, which is very rare [18].

ADA₁ catalyses the irreversible deamination of adenosine to inosine. Red Blood Cells (RBC) are in equilibrium with freely diffusing adenosine [19], pointing to an important role for this enzyme in the regulation of adenosine concentration.

Current interest has been focused on a wide variety of effects produced by adenosine via activation of cell surface adenosine receptors [20,21]. Adenosine counteracts insulin action in the liver by activating A2B receptors [22]. Adenosine seems to facilitate insulin action in adipocytes.

The adenosine deaminase complex protein [23] (ADPC) is identical with CD26, a T cell activating antigen and with a glycoprotein present in epithelial cells of various tissues. Recent data suggest that ADA₁ and CD26 are co-localized on the T cell surface but not inside cells.

Cells expressing ADA₁ and CD26 on the surface are much more resistant to the inhibitory effects of adenosine. These data suggest that ADA₁ on the cell surface is involved in an important immunoregulatory mechanism by which released ADA₁ binds to the cell surface of CD26, and this complex is capable of reducing the local concentration of adenosine [24].

In the present paper we have performed a more detailed analysis of the effect of ACP₁ polymorphism and have extended this study to the ADA₁ polymorphism. On the basis of the observations on women with repeated spontaneous miscarriages we would expect an optimal developmental context in zygotes carrying the ADA₁*2 and ACP₁*C alleles.

Methods

In the present study we examined 400 consecutive newborn infants from healthy puerperae. All infants were Caucasian from the population of Rome. Birth weight and

placental weight (wet, untrimmed) were registered in the delivery room. Gestational length was estimated from the date of the last menstrual period and checked against Dubowitz score as an additional index of neonatal maturity. Multiples were excluded. Mode of delivery does not alter the phenotype of enzyme considered.

The data presented in the paper were collected a few years ago and at that time there was not an established Ethical Committee. The project was discussed and approved in the Department. Informed verbal consent to participate in the study was obtained from the mothers. This has been recently (April 28, 2008) approved by the Institutional Ethical Committee.

Newborn blood samples were obtained from the placental side of the umbilical vein after umbilical cord section. The ACP₁ phenotype was determined in 361 newborns by starch gel electrophoresis on red blood cell hemolysates according to Harris and Hopkinson [25]. The acid phosphatase pattern is revealed by a solution of phenolphthalein diphosphate: the addition of ammonium solution reveals the area where phenolphthalein has been liberated in the areas of gel where ACP₁ activity is present. In European populations the presence of three common alleles *A, *B and *C determines the occurrence of six phenotypes: A, AB, B, AC, BC and C. Each of the homozygous A, B and C phenotypes are composed of two fractions, F and S, corresponding to fast and slow components of the electrophoretic pattern. Heterozygous phenotypes have a pattern corresponding to a mixture of homozygous types.

The ADA₁ phenotype was determined by starch gel electrophoresis on red blood cell hemolysates according to Spencer et al [26]. Inosine produced at the sites of ADA₁ activity is converted in hypoxanthine in the presence of nucleoside phosphorylase and phosphate. The hypoxanthine is then oxidized by the action of xanthine oxidase, and during this reaction the tetrazolium salt MTT is reduced in the presence of phenazine methosulphate to a blue insoluble formazan. In the ADA₁1 type there are three regularly spaced components which exhibit decreasing staining intensity in order of their anodal electrophoretic mobilities. In the ADA₁2 type there are also three isozymes and their relative intensities and relative electrophoretic mobilities are very similar to those of the ADA₁1 pattern. The difference between ADA₁1 and ADA₁2 is that the ADA₁2 pattern is appreciably slower than the ADA₁1 pattern. The pattern exhibiting four isozymes, designated ADA₁2/1, has the appearance of a mixture of ADA₁1 and ADA₁2 patterns.

In the last few years, in our laboratory, determination of ADA₁ and ACP₁ genotypes has been performed routinely

on DNA. In our laboratory the comparison of classical with DNA methods has shown practically no differences between phenotypic and genotypic classifications. On a sample of 50 subjects in which ACP₁ and ADA₁ phenotypes were determined by DNA and classical methods only one difference was observed for ACP₁ and no difference for ADA₁.

Correlation analysis was performed by SPSS programs. Differences between correlation coefficients were evaluated according to Snedecor and Cochran [27]. The distribution of ACP₁ phenotypes among newborns does not differ statistically from Hardy-Weinberg expectation.

Results

Table 1 shows demographic parameters of the sample study. Table 2 shows the distribution of ACP₁ phenotypes and developmental parameters for each phenotype. No statistical significant difference among ACP₁ phenotypes is observed for BW, PW and gestational duration. CA phenotype shows low values for all parameters, but these values are not statistically different from those of other ACP₁ phenotypes. BW-PW correlation analysis shows highly significant differences among ACP₁ phenotypes. The highest correlation coefficient is observed for CA phenotype and the lowest for B phenotype.

Figures 1, 2, 3, and 4 illustrate the relationship between BW-PW correlation and relevant ACP₁ parameters: F and S activity, F/S activity ratio and total activity. The BW-PW correlation is negatively associated with F concentration (Fig 1) and F/S ratio (Fig 3). A, CA and CB phenotypes that share a medium-low F activity and F/S ratio have a high BW-PW correlation, while the B phenotype, which has the highest F activity and F/S ratio, has a low BW-PW correlation. No association is observed for S isoform concentration (Fig 2) and ACP₁ total activity (Fig 4).

Table 3 shows the effect of smoking, maternal age, gestational age, parity and gender on the relationship between ACP1 and BW-PW correlation. For this analysis two classes of ACP1 phenotypes have been considered: A, CA, CB phenotypes with medium-low F activity, and B and BA phenotypes with medium-high F activity. In A, CA, CB

Table 1: Maternal and neonatal parameters in the sample study

	Mean	Proportion	S.E.
Maternal Age (yrs)	28.5		0.3
Gestational Age (wks)	39.6		0.12
Birth Weight (gr)	3269		29
Placental Weight (gr)	578		8
Smokers		38%	
Male Infant		54%	

Table 2: Distribution of neonatal parameters in relation to ACP₁ phenotypes.

	ACP ₁ phenotypes						Significance of difference among phenotypes (p)
	A	B	C	BA	CA	CB	
Absolute frequencies	34	163	2	123	9	30	
Percent frequencies	9.4%	45.2%	0.6%	34.1%	2.5%	8.3%	
<i>Birth weight (gr)</i>							
Mean	3345	3250	3645	3338	2837	3244	0.141 (*)
S.E.	85	46	575	44	333	90	
<i>Placental weight(gr)</i>							
Mean	590	594	615	571	517	540	0.491(*)
S.E.	23	17	135	12	46	21	
<i>Gestational age(wks)</i>							
Mean	40.00	39.61	39.50	39.77	38.11	39.53	0.405(*)
S.E.	0.41	0.20	1.50	0.15	1.57	0.40	
Median	40.00	40.00	39.50	40.00	40.00	40.00	
Correlation between birth weight and placental weight (r)	0.69	0.18	-	0.31	0.77	0.68	0.005(**)
Significance of r (p)	0.001	0.029	-	0.001	0.025	0.001	

Significance of differences among means (*) refers to Variance analysis. Significance of differences among correlation coefficients (**) has been calculated according to Snedecor and Cochran.

phenotypes only maternal age exhibits some effect on the BW-PW correlation, while among B and BA phenotypes most variables show highly significant effects on the BW-PW correlation.

Table 4 shows the distribution of ADA₁ phenotypes and developmental parameters for the ADA₁1 phenotype and for carriers of the ADA₁*2 allele. The BW-PW correlation coefficient is higher among carriers of the ADA₁*2 allele than among ADA₁1 phenotype carriers (p = 0.04). No significant difference between ADA₁ phenotypes has been observed for BW, PW and gestational duration.

Table 5 shows the effect of smoking, maternal age, gestational age, parity and gender on the relationship between ADA₁ and BW-PW correlation. Significant effects of these variables were observed in the correlations of BW-PW in both ADA₁ subjects and in carriers of the ADA₁*2 allele.

The BW-PW correlation is influenced by environmental and developmental variables, but as shown in tables 3 and 5 the differential effects of ACP₁ and ADA₁ phenotypes on this correlation do not appear in general to be significantly influenced by these variables.

Table 6 shows the correlation between BW and PW according to the joint ACP₁-ADA₁ phenotype. Both ADA₁ and ACP₁ phenotypes were determined in 327 infants. There is a highly significant difference among joint phenotypes (p = 0.000): the highest correlation is observed in subjects who carry A or CA or CB phenotypes and the ADA₁*2 allele, while the lowest correlation is observed when the ADA₁ 1 phenotype is associated with B or BA phenotypes.

Discussion and conclusion

The present data suggest that foetuses with low ADA₁ activity, associated with medium-low ACP₁ F isoform activity, have the best correlation between BW and PW, suggesting a most favourable situation for the development of the fetoplacental unit. These observations agree with those expected on the basis of previous data on women with repeated spontaneous miscarriages that demonstrated a cooperative protective effect of ADA₁*2 and ACP₁*C alleles against fetal loss. Thus, the two lines of evidence support the hypothesis that foetuses with low ADA₁ activity and low ACP₁ F isoform activity have a balanced development of fetoplacental unit and a higher probability of survival compared to other foetuses.

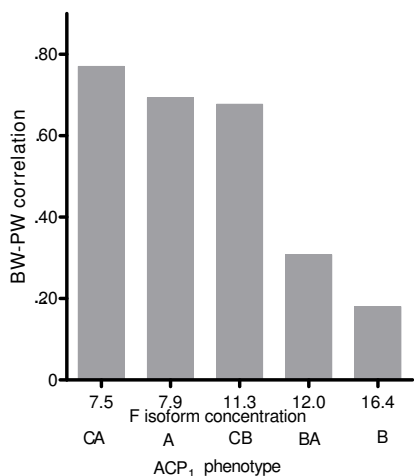


Figure 1
The relationship between F isoform concentration and BW-PW correlation. The term BW-PW correlation expresses the correlation between birth weight and placental weight. ACP1 is the acid phosphatase locus I. A, B, C, BA, CA, CB are the ACP₁ phenotypes. In abscissa F isoform concentrations of each ACP₁ phenotype are also reported. The rank correlation coefficient according to Spearman (27) between BW-PW correlation and F isoform concentration is $r_s = -1$, $p < 0.01$.

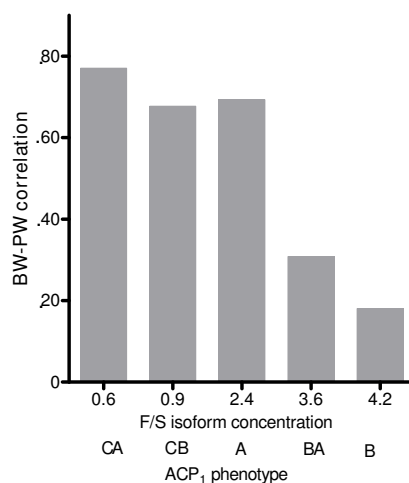


Figure 3
The relationship between F/S isoform concentration and BW-PW correlation. The term BW-PW correlation expresses the correlation between birth weight and placental weight. ACP1 is the acid phosphatase locus I. A, B, C, BA, CA, CB are the ACP₁ phenotypes. In abscissa F/S isoform concentrations of each ACP₁ phenotype are also reported. The rank correlation coefficient according to Spearman (27) between BW-PW correlation and F/S isoform concentration is $r_s = -0.9$, $p < 0.05$.

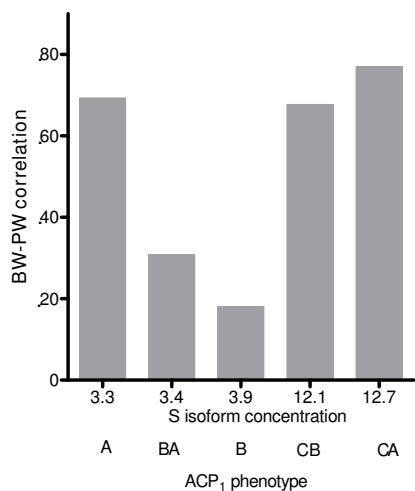


Figure 2
The relationship between S isoform concentration and BW-PW correlation. The term BW-PW correlation expresses the correlation between birth weight and placental weight. ACP1 is the acid phosphatase locus I. A, B, C, BA, CA, CB are the ACP₁ phenotypes. In abscissa S isoform concentrations of each ACP₁ phenotype are also reported. The rank correlation coefficient according to Spearman (27) between BW-PW correlation and S isoform concentration is $r_s = 0.3$, p not significant.

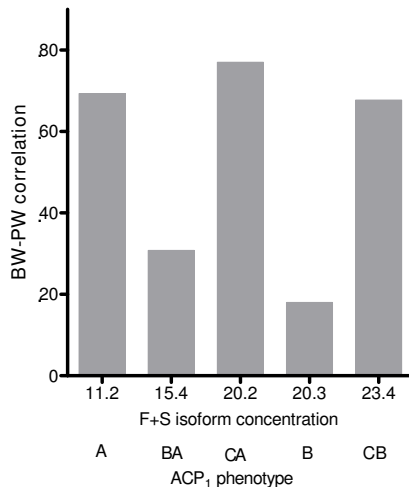


Figure 4
The relationship between F+S isoform concentration and BW-PW correlation. The term BW-PW correlation expresses the correlation between birth weight and placental weight. ACP1 is the acid phosphatase locus I. A, B, C, BA, CA, CB are the ACP₁ phenotypes. In abscissa F+S isoform concentrations of each ACP₁ phenotype are also reported. The rank correlation coefficient according to Spearman (27) between BW-PW correlation and F+S isoform concentration is $r_s = 0.3$, p not significant.

Table 3: Correlation between birth weight and placental weight in relation to ACP₁ F isoform activity.

Sample	ACP ₁ F isoform activity				
	Medium-high (B+BA)		Medium-low (A+CA+CB)		
	r	p*	r	p*	
All subjects	0.209	0.001	0.687	0.000	
<i>Smoking</i>					
	Yes	0.122	0.243	0.696	0.000
	no	0.241	0.002	0.710	0.000
	Significance of difference p**	0.000		0.750	
<i>Maternal age (yrs)</i>					
	≤28	0.027	0.745	0.723	0.000
	>28	0.478	0.000	0.640	0.000
	Significance of difference p**	0.000		0.025	
<i>Gestational age (wks)</i>					
	≤37	0.183	0.440	0.731	0.160
	>37	0.228	0.000	0.655	0.000
	Significance of difference p**	0.450		0.950	
<i>Birth order</i>					
	1	0.305	0.001	0.672	0.000
	≥ 2	0.158	0.060	0.703	0.000
	Significance of difference p**	0.000		0.350	
<i>Sex</i>					
	male	0.171	0.044	0.712	0.000
	female	0.301	0.001	0.655	0.000
	Significance of difference p**	0.000		0.170	

Significance of difference between correlation coefficients has been calculated according to Snedecor and Cochran. p* refers to significance of correlation coefficient. p** refers to significance of difference between the two classes. For "All subjects" the difference of correlation coefficients between (B+BA) vs (A+CA+CB) i.e. 0.209 vs 0.687 is highly significant: p = 0.002.

The exact mechanism underlying the statistical association of ADA₁ and ACP₁ with the BW-PW correlation is not known at present. An immunological mechanism is supported by the well known relationship between ADA₁ and immune diseases and between ACP₁ and T cell activation. A relative depression of T cell activation due to low level of F ACP₁ isoform and to higher concentration of adenosine (due to the low activity of ADA₁*2 carriers) could modulate the feto-maternal immunological relationship resulting in a balanced development of the two portions of feto-placental unit.

A metabolic mechanism may be operative in which ACP₁, acting as phosphotyrosine phosphatase, could have an

important role in the modulation of glycolytic rate through the control of insulin receptor activity and of band 3 protein phosphorylation status. Additionally, catalysing the conversion of flavin-mononucleotide (FMN) in riboflavin, the enzyme may influence flavo-enzyme activity and energy metabolism [7]. In turn, with respect to ADA₁ activity, recent studies have shown that adenosine counteracts insulin action in the liver by activating A2B receptors [20-22]. On the basis of these actions on glucose metabolism, ACP₁ F isoform activity coupled with low ADA₁ activity could have favourable effects on the development of the feto-placental unit.

Table 4: Parameters distribution of newborns in relation to ADA₁ phenotypes.

	ADA ₁ phenotypes		Significance of difference between phenotypes (p)
	ADA ₁ I	(ADA ₁ 2/I+ADA ₁ 2)	
Absolute frequencies	317	58	
Percent frequencies	84.5%	15.5%	
<i>Birth weight (gr)</i>			
Mean	3279	3311	0.612*
S.E.	32	54	
<i>Placental weight (gr)</i>			
Mean	584	545	0.103*
S.E.	10	13	
<i>Gestational age (wks)</i>			
Mean	39.63	39.84	0.505*
S.E.	0.13	0.24	
Median	40.00	40.00	
Correlation between birth weight and placental weight (r)	0.289	0.552	0.040**
Significance of r (p)	0.000	0.000	

Significance of difference between means (*) refers to Variance analysis. Significance of difference between correlation coefficients (**) has been calculated according to Snedecor and Cochran

Regarding the effect of genetic variability of ACP₁ on the correlation of BW with PW, it is interesting to speculate on the possible selective advantage of the *A and *C alleles over the *B allele. The *B allele is the most frequent in all human populations, and the *A allele is present with variable frequencies in all major ethnic groups, while *C allele is present with appreciable frequencies only in Caucasians.

Our data suggest that the optimal BW-PW correlation is seen in carriers of the ACP₁*C allele and in the homozygous A phenotype (table 4 and Fig 1), while the heterozygous BA phenotype shows an intermediate value between B and A (Fig 1). Interestingly, in A, CA and CB phenotypes the BW-PW correlation is hardly influenced by environmental circumstances, while in B and BA phenotypes the environmental variables exert considerable effects on this correlation. Thus, ACP₁*A and *C variants could have a selective advantage during intrauterine life on the fundamental ACP₁*B allele. This might have contributed to an increase in frequencies of the ACP₁*A and *C alleles to polymorphic values and could presently contribute to maintenance of the ACP₁ polymorphism in human populations.

Competing interests

The authors declare that they have no competing interests.

Table 5: Correlation between birth weight and placental weight in relation to ADA₁ phenotypes.

Sample	ADA ₁ phenotypes				
	ADA ₁ I		(ADA ₁ 2/I+ADA ₁ 2)		
	r	p*	r	p*	
All subjects	0.289	0.000	0.552	0.000	
Smoking					
	Yes	0.275	0.003	0.484	0.036
	no	0.288	0.000	0.589	0.000
	Significance of difference p**	0.300		0.150	
Maternal age (yrs)					
	≤28	0.104	0.181	0.679	0.000
	>28	0.559	0.000	0.341	0.120
	Significance of difference p**	0.000		0.000	
Gestational age (wks)					
	≤37	0.292	0.187	0.922	0.078
	>37	0.286	0.000	0.560	0.000
	Significance of difference p**	0.900		0.350	
Birth order					
	1	0.449	0.000	0.572	0.001
	≥ 2	0.212	0.009	0.543	0.003
	Significance of difference p**	0.000		0.700	
Sex					
	male	0.268	0.000	0.359	0.078
	females	0.344	0.000	0.692	0.000
	Significance of difference p**	0.000		0.000	

Significance of difference between correlation coefficients has been calculated according to Snedecor and Cochran. p* refers to significance of correlation coefficient. p** refers to significance of difference between the two classes. For "All subjects" the difference of correlation coefficients between ADA₁I and (ADA₁2/I+ADA₁2) i.e. 0.289 vs 0.552 is statistically significant: p = 0.04.

Table 6: Correlation between birth weight (BW) and placental weight (PW) according to the joint ACP₁-ADA₁ phenotype.

	Joint ACP ₁ -ADA ₁ phenotype			
	-	-	+	+
Presence of ADA*2 allele	-	-	+	+
Presence of A or CA or CB types	-	+	-	+
BW-PW correlation coefficients (r)	0.193	0.613	0.648	
Significance of r (p)	0.004	0.000	0.059	
Total n°	217	101	9	

Significance of difference among correlation coefficients p = 0.000. Significance of difference among correlation coefficients has been calculated according to Snedecor and Cochran

Authors' contributions

GBF, BE, MA and BA have been involved in the conception and design of the study, in drafting the manuscript and in its critical revision. GBF and BE have interpreted the data and performed the statistical analyses. PA and CL have contributed to the revision of the manuscript, to the acquisition of the data and to coordination of study. All authors have read and approved the final manuscript.

Acknowledgements

We wish to thank Prof. James MacMurray (Los Angeles, CA) for editing the manuscript.

References

- Nicotra M, Bottini N, La Torre M, Amante A, Bottini E, Gloria-Bottini F: **Repeated spontaneous abortion. Cooperative effects of ADA and ACPI genetic polymorphisms.** *Am J Reprod Immunol* 2007, **58**:1-10.
- Barker DJP, Bull AR, Osmond C, Simmonds SJ: **Fetal and placental size and risk of hypertension in adult life.** *BMJ* 1990, **301**:259-262.
- Edwards A, Megrens A, Peek M, Wallace EM: **Sexual origins of placental dysfunction.** *Lancet* 2000, **355**:203-204.
- Williams LA, Evans SF, Newnham JP: **Prospective cohort study of factors influencing the relative weights of the placenta and the newborn infant.** *BMJ* 1997, **314**:1864-1871.
- Sivarao S, Viyadaran MK, Jammal AB, Zainab S, Goh YM, Rames KN: **Weight, volume and surface area of placenta of normal pregnant women and their relation to maternal and neonatal parameters in Malay, Chinese and Indian ethnic groups.** *Placenta* 2002, **23**:691-696.
- Gloria-Bottini F, Bottini N, Cosmi E, Cosmi EV, Bottini E: **The effect of gender and ACPI genetic polymorphism on the correlation between birth weight and placental weight.** *Placenta* 2005, **26**:846-848.
- Bottini N, Bottini E, Gloria-Bottini F, Mustelin T: **Low-molecular-weight protein tyrosine phosphatase and human disease: in search of biochemical mechanisms.** *Arch Immunol Therap Exp* 2002, **50**:95-104.
- Spencer N, Hopkinson DA, Harris H: **Quantitative differences and gene dosage in the human red cell acid phosphatase polymorphism.** *Nature* 1964, **201**:299-300.
- Dissing J: **Human red cell acid phosphatase (ACPI) genetic, catalytic and molecular properties.** In *PhD thesis* Kobenhavn Universitat, Kobenhavn, Denmark; 1993.
- Dissing J: **Immunochemical characterization of human red cell acid phosphatase isozymes.** *Biochem Genet* 1987, **25**:901-917.
- Mansfield E, Sensabaugh GF: **Red cell acid phosphatase: modulation of activity by purines.** In *The red cell Volume 21*. Edited by: Brewer GJ, Allan R. Liss. New York; 1978:233-247.
- Boivin P, Galand C: **The human red cell acid phosphatase is a phosphotyrosine protein phosphatase which dephosphorylates the membrane protein band 3.** *Biochem Biophys Res Commun* 1986, **134**:557-564.
- Fuchs KR, Shekels LL, Bernlohr DA: **Analysis of the ACP I gene product-classification as an FMN phosphatase.** *Biochem Biophys Res Commun* 1992, **189**:1598-1605.
- Gloria-Bottini F, Gerlini G, Lucarini N, Borgiani P, Amante A, La Torre M, Antonacci E, Bottini E: **Phosphotyrosine protein phosphatases and diabetic pregnancy. An association between low molecular weight acid phosphatase (ACPI) and degree of glycemic control.** *Experientia* 1996, **52**:340-343.
- Lucarini N, Antonacci E, Bottini N, Borgiani P, Faggioni G, Gloria-Bottini F: **Phosphotyrosine-protein phosphatase and diabetic disorders. Further studies on the relationship between low molecular weight acid phosphatase genotype and degree of glycemic control.** *Disease Markers* 1998, **14**:121-125.
- Bottini N, Stefanini L, Williams S, Alonso A, Jascur T, Abraham RT, Couture C, Mustelin T: **Activation of ZAP-70 through specific dephosphorylation at the inhibitory Tyr-292 by the low molecular weight phosphotyrosine phosphatase (LMPTP).** *J Biol Chem* 2002, **277**:24220-24224.
- Spencer N, Hopkinson D, Harris H: **Adenosine deaminase polymorphism in man.** *Ann Hum Genet* 1968, **32**:9-14.
- Battistuzzi C, Scozzari R, Santolamazza P, Terrenato L, Modiano G: **Comparative activity of red cell adenosine deaminase allelic forms.** *Nature* 1974, **251**:712.
- McKusik VA: **Mendelian Inheritance in Man.** 11th edition. Baltimore, MD, The Johns Hopkins University Press; 1994.
- Xu B, Berkich DA, Crist Gh, LaNoue KF: **A₁ adenosine receptor antagonism improves glucose tolerance in Zucker rats.** *Am J Physiol* 1998, **274**:E271-279.
- Richardson PJ: **Blocking adenosine with antisense.** *Nature* 1997, **385**:684-685.
- Yasuda N, Inoue T, Horizoe T, Nagata K, Minami H, Kawata T, Hoshino Y, Harada H, Yoshikawa S, Asano O, Nagaoka J, Murakami M, Abe S, Kobayashi S, Tanaka I: **Functional characterization of the adenosine receptor contributing to glycogenolysis and gluconeogenesis in rat hepatocytes.** *Eur J Pharmacol* 2003, **459**:159-166.
- Kameoka J, Tanaka T, Nojima Y, Schlossman SF, Morimoto C: **Direct association of adenosine deaminase with T cell activation antigen CD26.** *Science* 1993, **261**:466-469.
- Dong RP, Kameoka J, Hegen M, Tanaka T, Xu Y, Schlossman SF, Morimoto C: **Characterization of adenosine deaminase binding to human CD26 on T cells and its biologic role in immune response.** *J Immunol* 1996, **156**:1349-1355.
- Harris H, Hopkinson DA: **Handbook of enzyme electrophoresis in human genetics.** Amsterdam: Elsevier; 1976.
- Spencer N, Hopkinson D, Harris H: **Adenosine deaminase polymorphism in man.** *Ann Hum Genet* 1968, **32**:9-14.
- Snedecor GW, Cochran WG: **Statistical methods.** Ames: Iowa State University; 1989.

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